



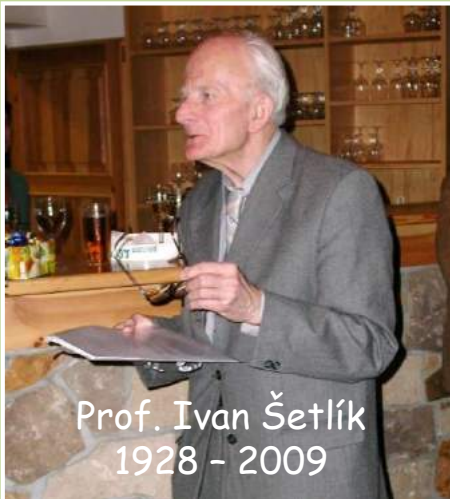
Centre ALGATECH
Institute of Microbiology
Třeboň, Czech Republic

Microalgal biotechnology in the Centre Algatech in Trebon. Overview of mass cultivation and downstreaming

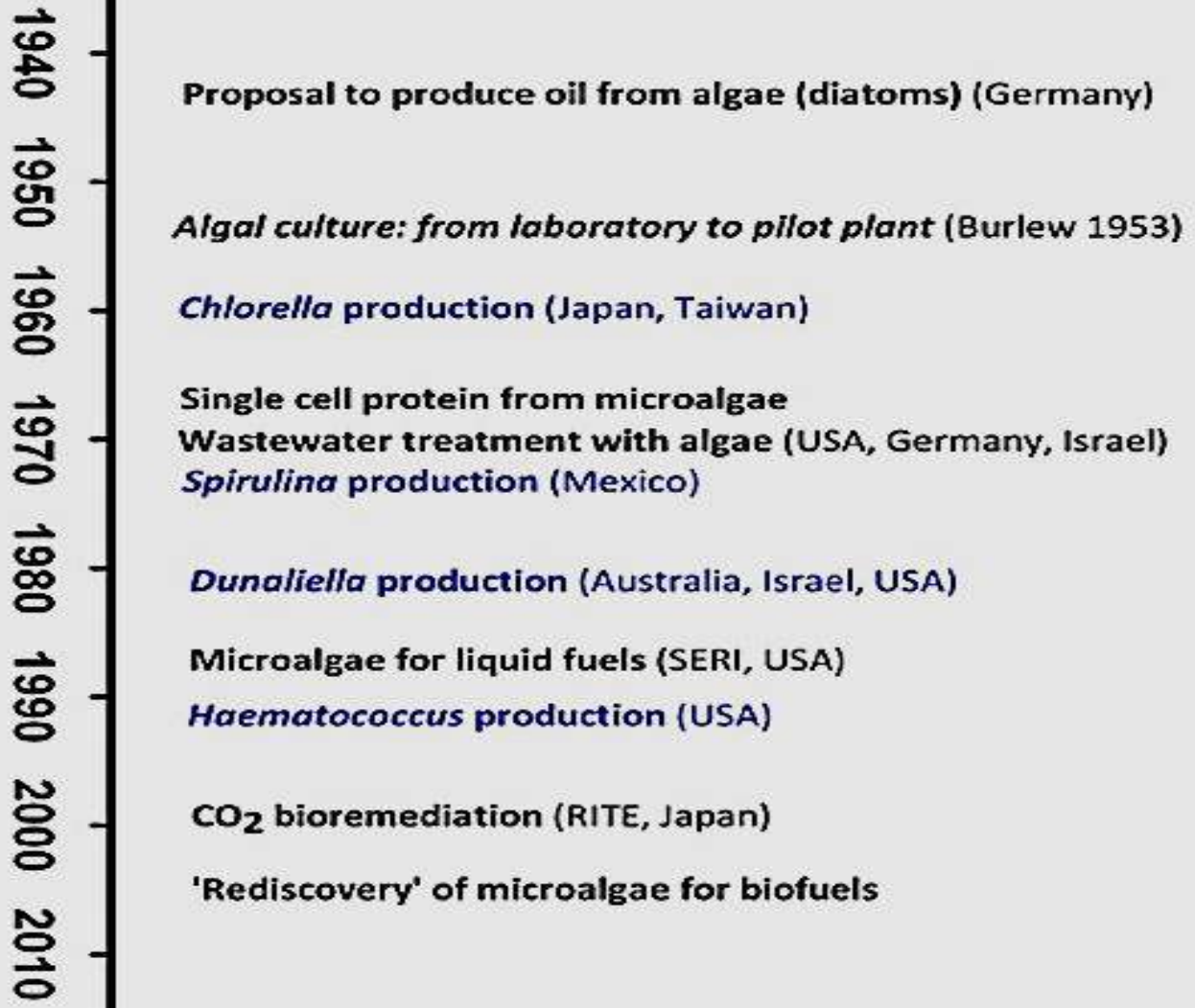
David Kubáč

Past and present: a timeline Microalgal Biotechnology in Czechoslovakia (Czech Republic)

Large scale cultivation
of microalgae
Institute of Microbiology
Třeboň, Czechoslovakia



Prof. Ivan Šetlík
1928 - 2009



1960s Inclined platforms with baffles - first large-scale system in Europe

Unique thin-layer cultivation system - highly productive, high cell density cultures designed in the early 1960s by I. Šetlík & co-workers for growth of microalgae
Institute of Microbiology, Academy of Science, Třeboň, 900 m²



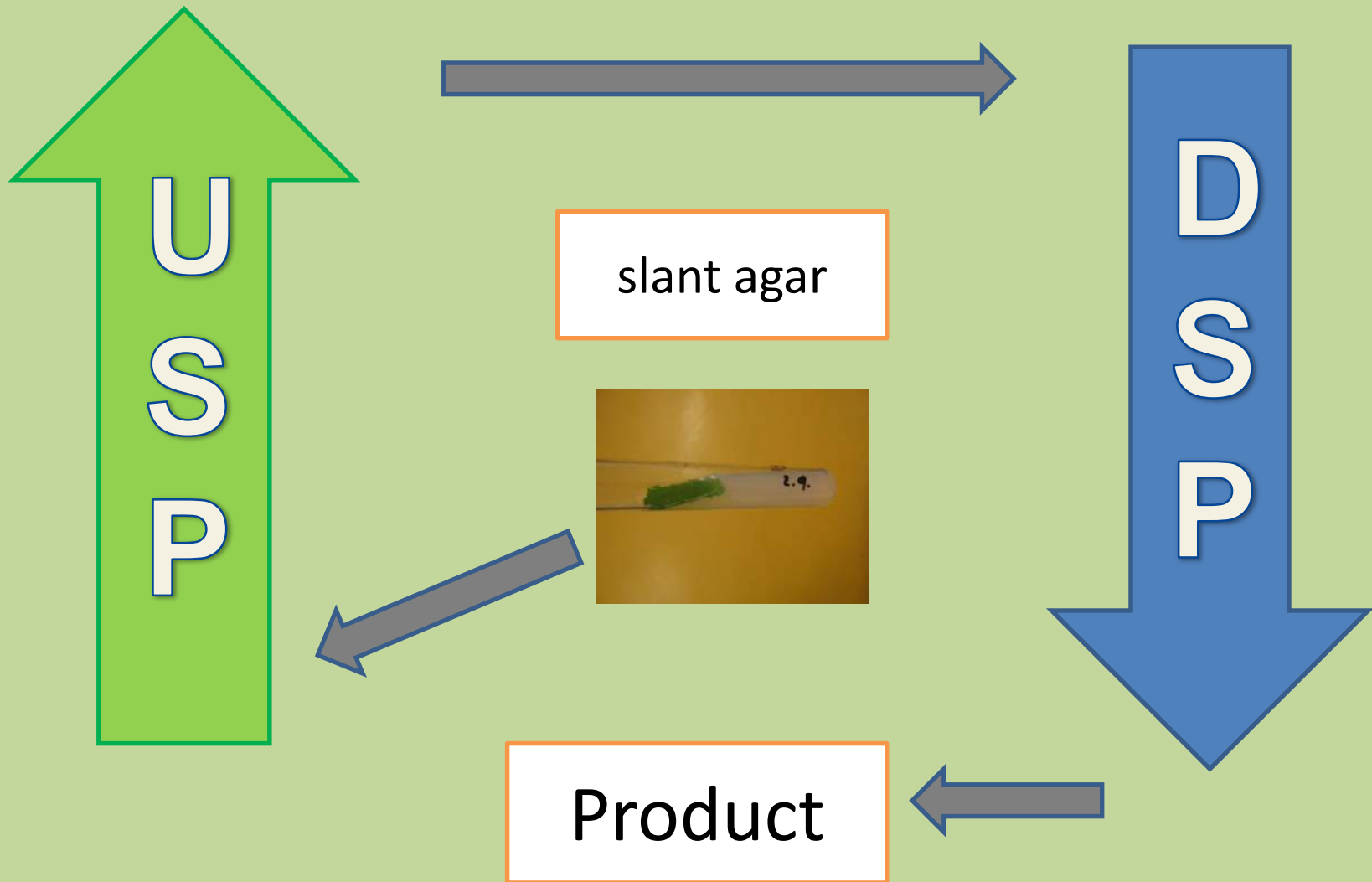
Šetlík et al. (1970) Dual purpose open cultivation units for large scale culture of algae in temperate zones. *Algological Studies* 1: 111-164.

Research topics

Laboratory of Algal Biotechnology

- Screening and selection of microalgae strains
- Design and construction of various cultivation units
- Optimisation of culturing regimes for microalgae
- Heterotrophic cultivation of microalgae
- Production of biomass as food and feed additives, for isolation of valuable compounds, etc.
- Identification and characterisation of bioactive compounds with potential pharmacological use – analytical techniques

Process



Advantages of heterotrophy

- controlled cultivation conditions
 - no weather influence
 - little space necessity
 - high yields
 - simple scale-up
-
- ability to direct produce at pharma grade
 - closed systems for safe GMO cultivation

Disadvantages heterotrophy

- limited group of strains
- ability vs. interest
- easy contamination with other heterotrophs
- few axenic strains in culture collections
- difficult screening
- carbon footprint cultivations

USP

- lab inoculum preparation
- F150 inoculum cultivation
- F1500 inoculation
- F1500 production batch cultivation



18 L



150 L



1500 L

DSP

- Biomass harvesting and dilution
- Centrifugation
- Disintegration
- Spray drying



Counter current Chromatography (CCC)

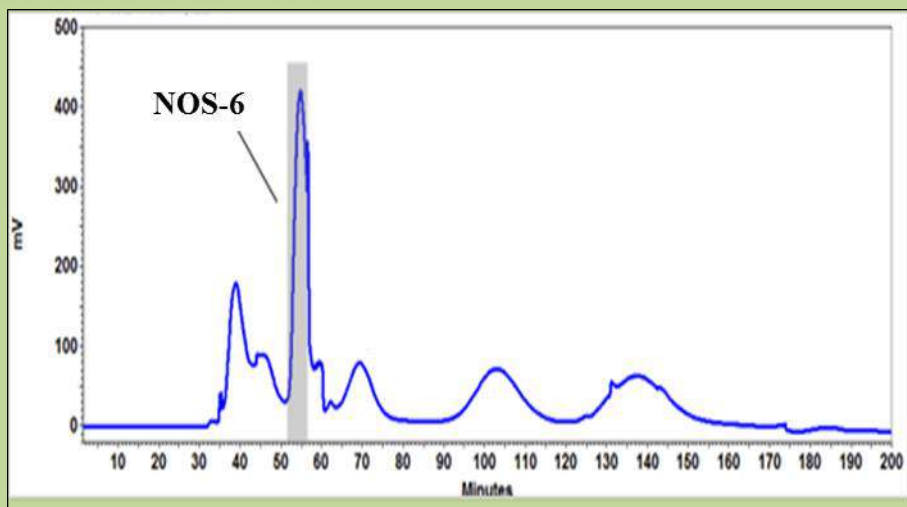
- CCC - liquid/liquid chromatography technique (two immiscible liquids)
- One liquid phase (the stationary phase) is retained in the column by centrifugal force, second (the mobile phase) is pumped through the column.

Benefits of CCC

- ✓ No risk of irreversible adsorption of analytes (no solid support).
- ✓ High loading capacity of sample (high throughput).
- ✓ Cost-effective technique (low solvent consumption).
- ✓ Greener technology (non-toxic and biodegradable solvents).
- ✓ Scalable - from laboratory to pilot and industrial size.
- ✓ Automated operation and fully predictable separation.
- ✓ High recovery of injected sample (liquid stationary phase).
- ✓ Multifunctional technique (extraction, fractionation, isolation, purification).

HPCCC separation of cold acetone-treated extract of *Nostoc* sp.20

HPCCC parameters	
Column volume	134 mL (Dynamic Extractions - Spectrum HPCCC)
Solvent System	<i>n</i> -hexane–ethyl acetate–MeOH–water (HEMWat, 4:5:4:5, v/v/v/v)
Rotational speed	1000 rpm
Flow rate	1 mL/min
Stationary phase	83%
Run Mode	Reverse, Isocratic
Injection Volume	3 mL
Sample	100 mg acetone-treated crude extract in 3 ml (UP/LP, 1:1)
Detection	280 nm



Scale-up

Traditional way



Test the viability of the big process in lab scale.

Cultivation *J. marinum*

Inoculation: 16-18L dense culture

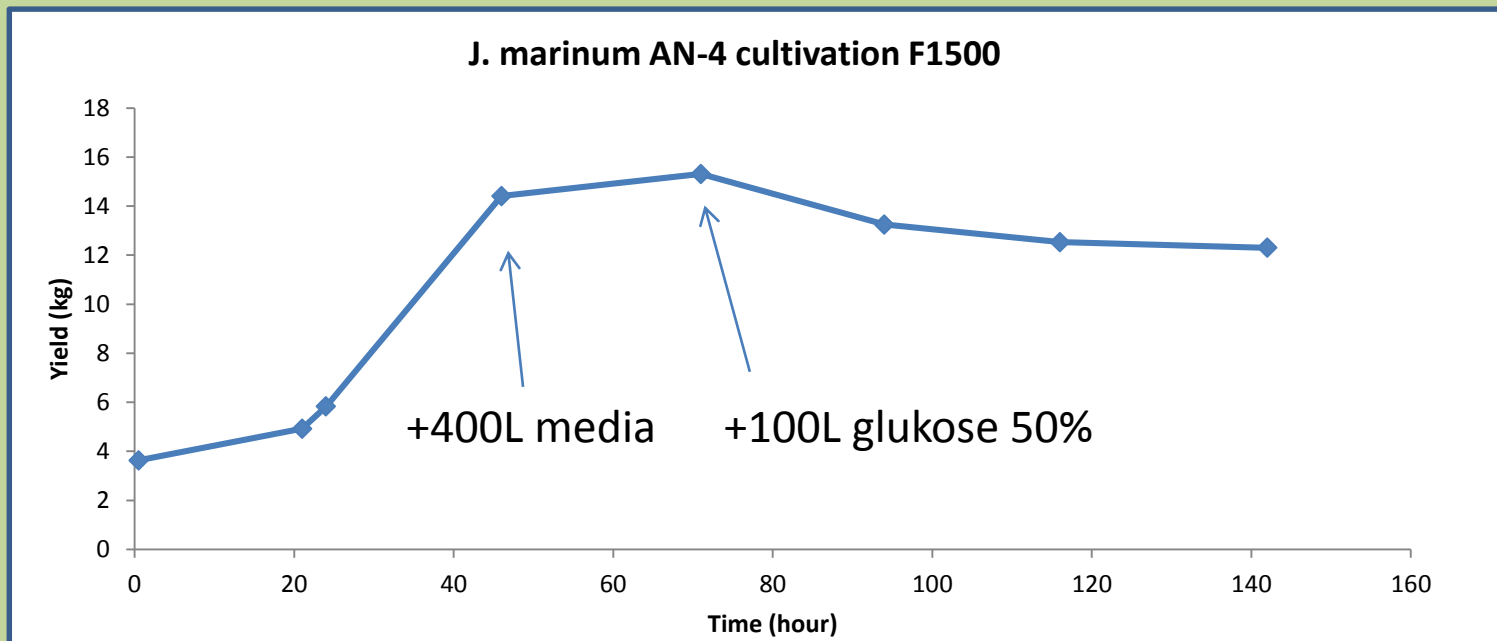
Cultivation:

two steps: 1. grow

27 °C

2. lipid production

23-24 °C

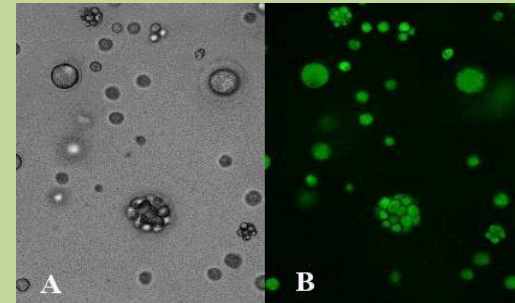


DSP

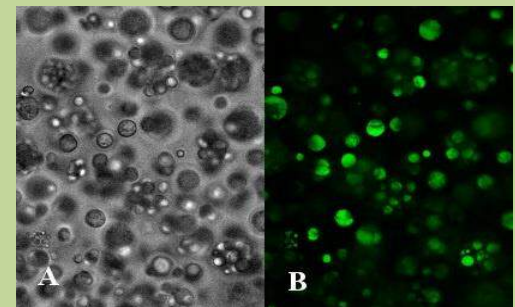
Lamellar centrifuge EVODOS 10

- fixed rpm 1400
- feed regulation 700L/hour

separation efficiency 51%



End sample F1500



Spray dried sample F1500



Lipid
 extract

Summary results

Batch	FA (g/kg DW)	DHA mg/L per day
1.	33.7	60
2.	108.8	205
3.	173.64	260

Vessel F1500 before harvesting

Day	FA (g/kg)	DHA (mg/kg DW)
4	87.21	53.07
5	104.07	59.07
6	173.64	92.02

Acknowledgement

Jiří Kopecký	Algatech
Jiří Masojídek	Algatech
Jose Cheel Horna	Algatech
Jan Hiterholzinger	Algatech
Standa Pumpr	Algatech
Soňa Pekařová	Algatech
Jindřiška Paichlová	Algatech

Tomáš Humhal	ICT Prague
Dana Savická	ICT Prague

Petr Kaštánek	EcoFuel
Olga Kronusová	EcoFuel

Funding

- Ministry of Education, Youth and Sports
- Czech Academy of Science
- Czech Science Foundation



J. marinum AN4 spray dried



Chlorella vulgaris BEIJ/H14



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Thank you for attention